Nanion Technologies

Small Currents – Big Potential.

Electrophysiology for transporters using the "SURFE²R" technology

Dr. Maria Barthmes

Application Scientist & Product Manager SURFE²R Family Nanion Technologies



Nanion Technologies introduced



- University spin-off in 2002
- Instrumentation for research, drug screening and safety testing
- One of the leading providers of automated patch clamp systems

- Headquarters located in Munich, Germany
- ~100 employees worldwide
- Offices in USA, Japan, China



Electrophysiology for transporters

Many transporters generate transmembrane currents

- P-Type and F/V-Type ATPases
- Many SLC transporters (coupled transport)

Electrophysiology

- → No labels
- → Real time measurement
- → High information content
- → High experimental freedom

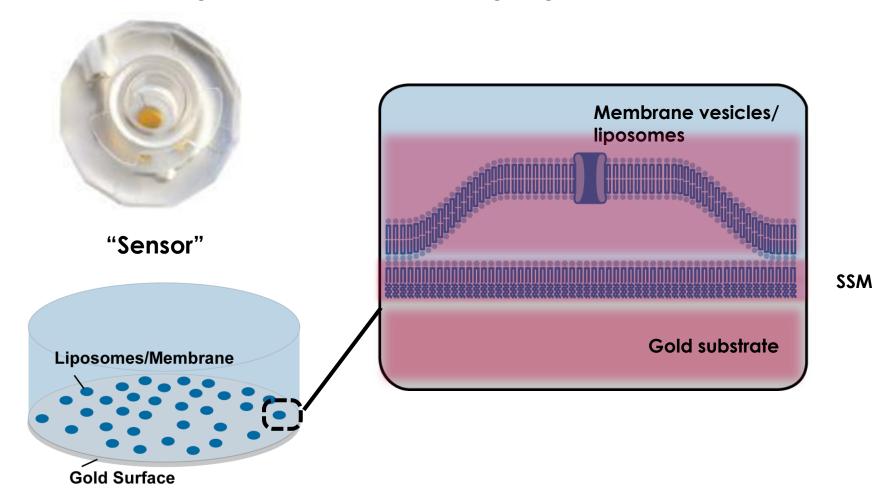
Patch Clamp/Voltage Clamp → low signal amplitude

Oocyte TEVC→ low throughput, compound concentrations

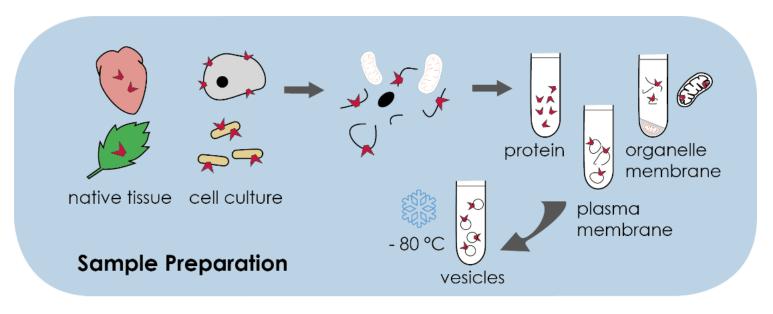
SSM-based Electrophyiology → invented for transporters

Solid supported membrane (SSM)-based electrophysiology

high amount of protein = high signal



The sample on the SSM



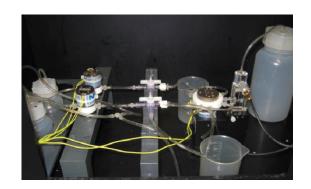
10 µl per 96 well plate ~ 0.1 µg protein/well



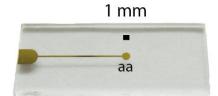


Insertion of the Sensor into a recording device

Custom built







SURFE²R N1





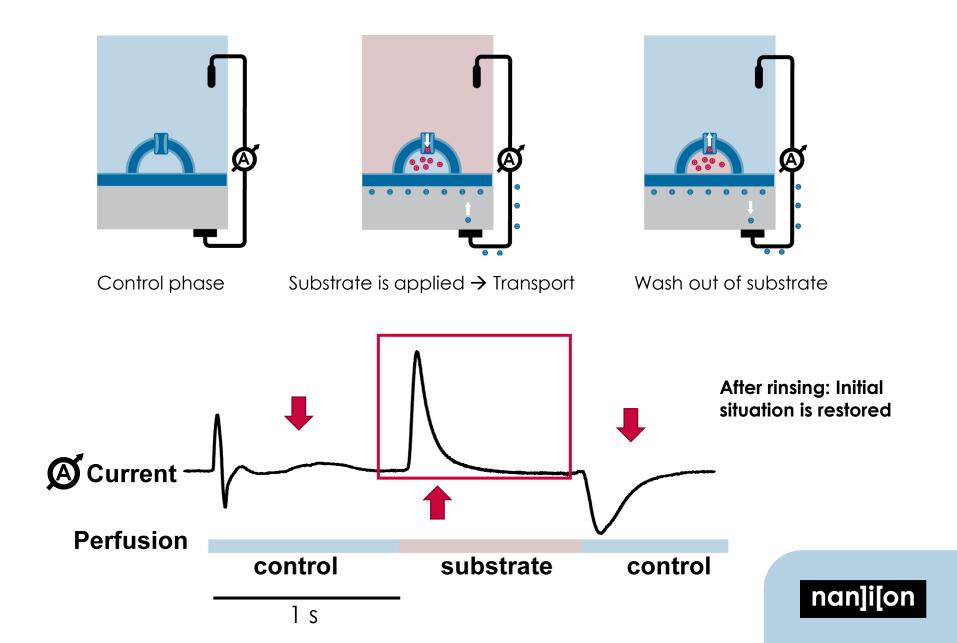


SURFE²R 96SE

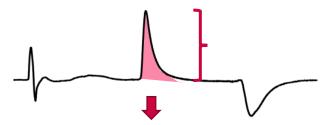




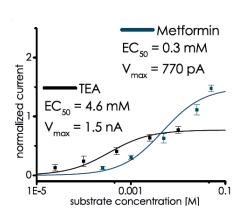
Activation by perfusion of the sensor



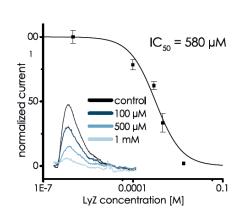
Experimental scope



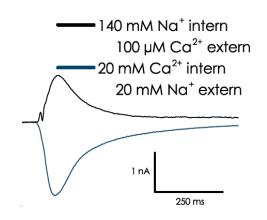
Substrates



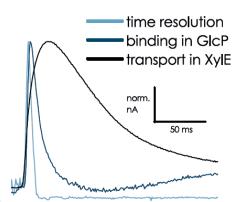
Inhibition



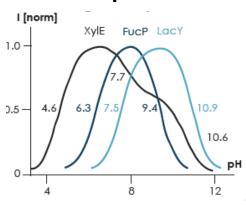
Environment



Mechanisms

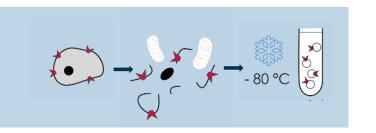


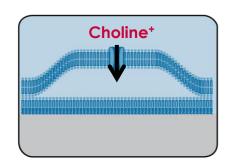
Comparison

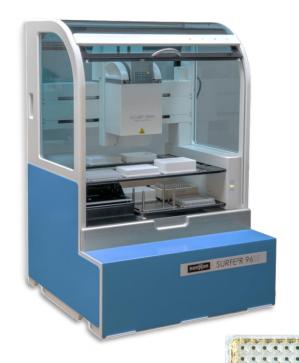


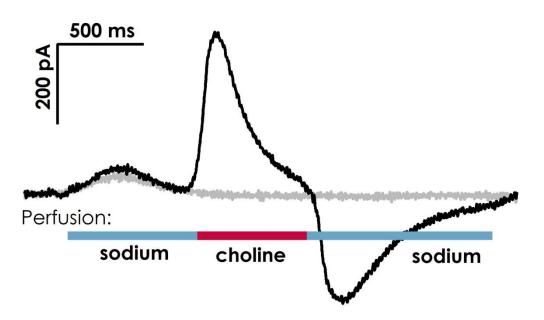
OCT2: Development and validation of an assay







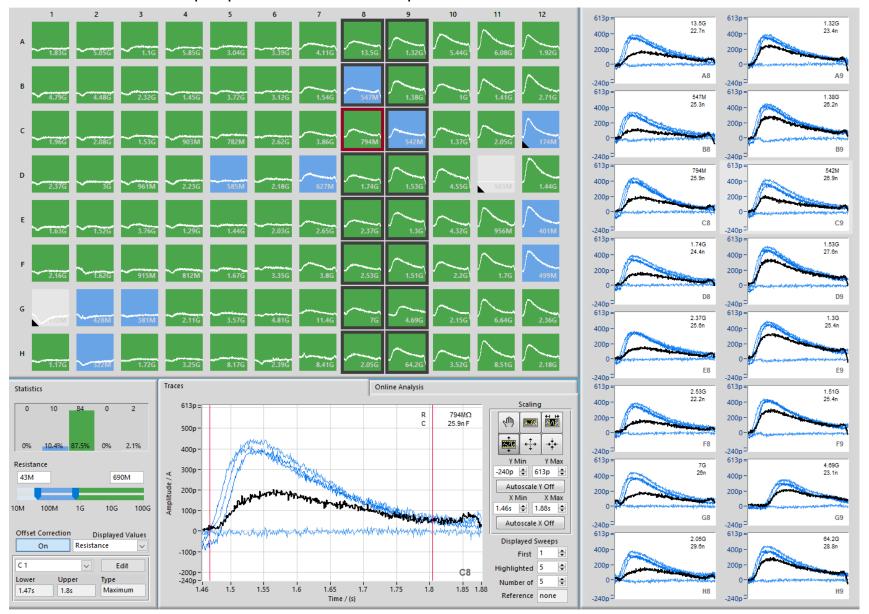




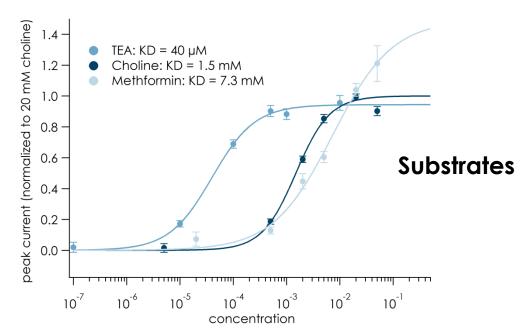


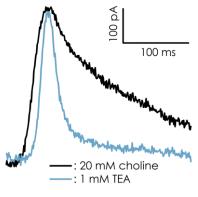
OCT2: Development and validation of an assay

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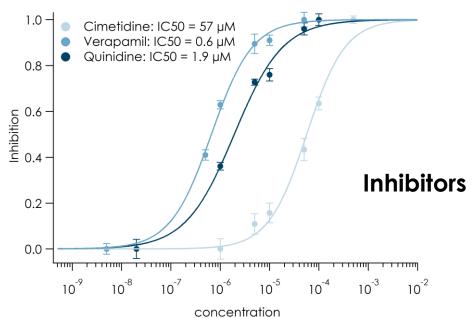


OCT2: Development and validation of an assay





	SSM	Published
TEA	40 µM	33-270 µM
Choline	1.5 mM	0.46 mM
Metformin	7.3 mM	0.6 -3.3 mM



	SSM	Published
Cimetidine	57 µM	8.6-120 µM
Verapamil	0.6 μΜ	13.4-85 µM
Quinidine	1.9 µM	8-87 µM



Other validated targets on the SURFE²R

<u>ATPases</u>
NaK-ATPase
HK-ATPase
SERCA
V-ATPase
FoF1-ATPase
Kdp-ATPase
CopA
ATP7A/B
VrPPase

Channels and Pores

Gramicidine

P2X2 nAChR A/M2

UCP1 (Slc25a7)

TRPC5
TRPA1
CFTR
AQP6

Redox-driven ion pumps

Complex I

MntH2

respiratory chain complex I/III

respiratory chain complex II/III

cyctochrome c-oxidase

respiratory chain complexes I/III/V

<u>Light-driven ion pumps</u>

Bacteriorhodopsin (BR)

Oxyrrhis marina Rhodopsin

Rhodopsin-2 (KR2)

Halorhodopsin (HR) Acerhodopsin

Characalda ala asia (Ci

<u>Pumps</u>

Channe	Irhoc	lopsin ((ChR)
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Exchangers

Inorganic ions	Amino acids	Sugars		Organic ions
NhaA	PEPT1 (Slc15a1)	SGLT1/2 (Slc5a1/a2	2)	OCT2 (Slc22a2)
NhaP	YdgR	MelB		CNT1 (Slc28a1)
NhaB	YhiP/DtpB	LacY		ANT (Slc25a4)
NCX1 (Slc8a1)	PutP	FucP		AAC
Clc-7	GltP	XylE		GAT1 (Slc6a1)
EcClc	EAAC1 (Slc1a1)	GlcP		BetP
NirC	PAT1 (Slc36a1)			CHT (Slc5a7)
Amt1-3	ArcD			NupC
AmtB	CAT2B (Slc7a2)			NacT (Slc13a5)
SulP	GlyT1/2 (Slc6a9/a5)			
NIS (Slc5a5)			<u>Uniporter</u>	<u>s</u>
NaPi2b (Slc34a2)		<u>Transporters</u>	Symporte	ers ers